



## Endophytic fungi in economically important plants: ecological aspects, diversity and potential biotechnological applications

*Fungos endofíticos em plantas economicamente importantes: aspectos ecológicos, diversidade e potenciais aplicações biotecnológicas*

<sup>1</sup>  Beatriz dos Santos SOUZA e <sup>2,\*</sup>  Taides Tavares dos SANTOS

1 Universidade Federal do Oeste do Pará (UFOPA), Instituto de Ciências e Tecnologia das Águas (ICTA), Campus de Santarém, Avenida Mendonça Furtado, n.º 2946, Fátima, 68040-470. Santarém – PA, Brasil.

2 Universidade Federal do Tocantins (UFT), Campus de Araguaína, Av. Paraguai, esq. c/ Rua Uxiramas, s/nº, Setor Cimba, 77.824-838. Araguaína – TO, Brasil.

\*Corresponding author: taides.tavares@hotmail.com

### OPEN ACCESS

### Additional informations

Received: 09/20/2016

Accept: 05/17/2017

Published: 08/07/2017

Editor

Claudio Alberto Gellis de Mattos  
Dias  
Instituto Federal do Amapá,  
Macapá, AP.

### Reviews process

Prot. 1212016R01 (Brasil)

Prot. 1212017R02 (Brasil)



JBFS all rights

Copyright: © 2017

### ABSTRACT

The aim of the present study was to perform a review of the literature to provide a brief overview of the ecological aspects, diversity and potential biotechnological applications of endophytic fungi associated with economically important plants, such as the common bean (*Phaseolus vulgaris*), cocoa (*Theobroma cacao*) and soybean (*Glycine max*). A high diversity of fungi has been reported on the leaves, stems and other organs of such plants. Studies on this subject are of considerable importance and directly contribute to the knowledge and conservation of fungal biodiversity, especially in Neotropical areas. Furthermore, such studies can open the door to a wide range of potential uses and/or modulations of fungal microbiota with the aim of enhancing the health and productivity of plants in agricultural practices, such as promoting plant growth or the biological control of diseases caused by phytopathogenic microorganisms. These endophytic fungi also represent a valuable source for prospecting secondary metabolites for biotechnological purposes.

**Keywords:** Biotechnology. Biological control. Symbiotic interactions. Secondary metabolites. Plant growth promotion

### RESUMO

O presente estudo, que consiste em uma revisão de literatura, teve o objetivo de apresentar um breve panorama acerca de aspectos ecológicos, diversidade e potenciais aplicações biotecnológicas de fungos endofíticos associados a plantas economicamente importantes, tais como o cacau (*Theobroma cacao*), o feijão comum (*Phaseolus vulgaris*) e a soja (*Glycine max*). Uma alta diversidade de fungos tem sido revelada em associação com folhas, caules e outros órgãos dessas plantas. Estudos com essa temática são de grande importância, uma vez que eles contribuem diretamente para o conhecimento e conservação da biodiversidade fúngica, sobretudo em áreas neotropicais. Além disso, podem abrir portas para uma vasta gama de potenciais usos e/ou modulações da microbiota fúngica associada a fim de incrementar a sanidade e, conseqüentemente, a produtividade do hospedeiro em práticas agrícolas, tais como a promoção do crescimento vegetal ou o controle biológico de doenças causadas por micro-organismos fitopatogênicos. Fungos endofíticos de plantas economicamente importantes podem, ainda, representar uma valiosa fonte para prospecção de metabólitos secundários de interesse biotecnológico.

**Palavras-chaves:** Biotecnologia. Controle biológico. Interação simbiótica. Metabólitos secundários. Promoção de crescimento vegetal.

## INTRODUCTION

Endophytic fungi are microorganisms that live in plant tissues for at least a part of their life cycle, apparently without causing any harm to the host (PETRINI, 1991; CABRAL et al., 1993). In recent years, considerable attention has been given to these fungi, with the conduction of a large number of studies involving a variety of approaches (AZEVEDO et al., 2000; HYDE; SOYTONG, 2008; KUMAR; KAUSHIK, 2012; NISA et al., 2015). The most commonly discussed issues regard the clarification of aspects related to the symbiosis between endophytic fungi and host plants (ARNOLD et al., 2001; FERREIRA et al., 2015), the composition of fungal communities associated with plant species in both temperate and tropical regions (GONZAGA et al., 2014; FERNANDES et al., 2015; SANTOS et al., 2016) and the functions of these microorganisms that favor the host, such as protection against pathogens (CLARKE et al., 2006; HANADA et al., 2008; GAO et al., 2010) and the promotion of plant growth (CLARKE et al., 2006; HANADA et al., 2008; Gao et al., 2010; OWNLEY et al., 2010). Focus has also been on the exploration of secondary metabolites of fungi to obtain new bioactive molecules with potential applications in the medical, pharmaceutical, industrial, agricultural and environmental fields (BUDHIRAJA et al., 2013; CAMPOS et al., 2015; CHEN et al., 2015; KUMAR et al., 2016).

Endophytic microorganisms are ubiquitous, having been encountered in many plant species growing under natural conditions (RODRIGUEZ et al., 2009; NISA et al., 2015). Endophytic fungi, in particular, have been investigated in a variety of plants ranging from grasses (MÜLLER; KRAUSS, 2005; REKHA; SHIVANA, 2014) to trees (Arnold et al., 2003; Campos et al., 2015). Studies have been conducted on medical plants (MUSSI-DIAS et al., 2012; FERREIRA et al., 2015), toxic plants (SOUZA et al., 2004; CAFÉU et al., 2005) and plants with recognized economic importance in Brazil and other countries, such as the common bean (GONZAGA et al., 2014; SANTOS et al., 2016), cocoa (MEJÍA et al., 2008; HANADA et al., 2010) and soybean (LEITE et al., 2013; Fernandes et al., 2015), which are responsible for the movement of large sums of money every year through local marketing and exportation. Moreover, many economically important plants are cultivated over large areas and are often affected by phytopathogenic fungi, pests and other organisms, which can result in decreased productivity. Thus, the study of endophytic fungi living in these plants can provide insights into the mediation of these and other common problems in agricultural production systems.

The aim of the present review was to provide a brief overview of the ecological aspects, diversity and potential biotechnological applications of endophytic fungi associated with economically important plants.

## ENDOPHYTIC FUNGAL DIVERSITY IN ECONOMICALLY IMPORTANT PLANTS

Some factors exert a qualitative and quantitative influence on the biodiversity of the endophytic fungal community of a plant, such as the age of the plant, the tissue or organ studied and methods employed for the isolation of fungi (PEIXOTO NETO et al., 2002; HYDE; SOYTONG, 2008). The endophytic fungal community inhabits different plant structures, such as leaves, petioles, reproductive structures, twigs, bark and roots (RODRIGUEZ *et al.*, 2009). A wealth of such fungi (most belonging to the phylum Ascomycota) has been detected (HYDE; SOYTONG, 2008). As summarized in Table 1, endophytic fungi have been investigated in different organs of a wide variety of plant species of economic interest, including food crops such as the common bean (*Phaseolus vulgaris* L.), cocoa (*Theobroma cacao* L.), soybean (*Glycine max* (L.) Merr.) and wheat (*Triticum aestivum* L.). A high diversity of fungi has been reported in

association with the leaves, stems and other organs of such plants. Studies on this subject are of considerable importance and directly contribute to the knowledge and conservation of fungal biodiversity, especially in Neotropical areas.

**Table 1.** Host plants, isolation parts, and frequent taxonomic groups of endophytic fungi isolated from economically important plants.

| Host plants  |   | Isolation parts                  | Frequent taxonomic groups  | References                     |
|--------------|---|----------------------------------|--|--------------------------------|
| Common names | Scientific names  |                                  |  |                                |
| Banana       | <i>Musa acuminata</i><br>Colla                              | leaves                           | <i>Xylaria</i> sp., <i>Colletotrichum musae</i> and <i>Cordana musae</i>               | Pereira; Vieira; Azevedo, 1999 |
| Cocoa        | <i>Theobroma cacao</i><br>L.                                | branches                         | <i>Fusarium</i> spp.   | Rubini et al., 2005            |
|              |   | stems and pods                   | <i>Coprinellus</i> sp.   | Crozier et al., 2006           |
|              |   | leaves and fruits                | (*1)   | Mejía et al., 2008             |
|              |   | branches, twigs and stem bark    | <i>Trichoderma</i> spp., <i>Pestalotiopsis</i> spp. and <i>Fusarium</i> spp.           | Hanada et al., 2010            |
| Coffee       | <i>Coffea arabica</i> L.                                    | leaves                           | <i>Colletotrichum</i> , <i>Xylaria</i> and <i>Guignardia</i>                           | Santamaría; Bayman, 2005       |
|              |   | leaves                           | (*1)   | Fernandes et al., 2009         |
|              | <i>C. arabica</i> L. and other coffee species               | leaves, roots, stems and berries | <i>Colletotrichum</i> , <i>Fusarium</i> , <i>Penicillium</i> and <i>Xylariaceae</i>    | Vega et al., 2010              |
| Common bean  | <i>Phaseolus vulgaris</i> L.                                | leaves                           | <i>Colletotrichum</i> , <i>Hannaella</i> , <i>Cochliobolus</i> and <i>Phomopsis</i> .  | Gonzaga et al., 2014           |
| Cowpea       | <i>Vigna unguiculata</i> (L.) Walp.                         | seeds                            | <i>Aspergillus</i> spp., <i>Penicillium</i> spp. and <i>Fusarium</i> spp.              | Rodrigues; Menezes, 2002       |
| Cupuassu     | <i>Theobroma grandiflorum</i> (Willd. ex Spreng.) K. Schum. | branches, twigs and stem bark    | <i>Pestalotiopsis</i> spp.   | Hanada et al., 2010            |
| Grapevine    | <i>Vitis vinifera</i> L.                                    | stems                            | <i>Alternaria</i> sp., <i>Epicoccum nigrum</i> and <i>Aureobasidium pullulans</i>      | Pancher et al., 2012           |
| Maize        | <i>Zea mays</i> L.  | leaves and stems                 | <i>Alternaria alternata</i> and <i>Aureobasidium pullulans</i> var. <i>melanigerum</i> | Fisher et al, 1992             |
|              |   | grains and roots                 | (*1)   | Orole; Adejumo, 2011           |

To be continued

Table 1. Continued

| Host plants  |                                  | Isolation parts                     | Frequent taxonomic groups   | References                                  |
|--------------|----------------------------------|-------------------------------------|---|---|
| Common names | Scientific names                 |                                     |   |   |
| Orange       | <i>Citrus</i> spp.               | leaves and seeds                    | <i>Colletotrichum gloeosporioides</i> ,<br><i>Guignardia citricarpa</i> and<br><i>Cladosporium</i> sp.  | Araújo et al., 2001                         |
| Pejibaye     | <i>Bactris gasipaes</i><br>Kunth | shoot tips                          | <i>Fusarium</i> spp., <i>Neotyphodium</i> sp.,<br><i>Epicoccum nigrum</i> ; <i>Colletotrichum</i> sp.<br>and <i>Alternaria gaisen</i>   | Almeida; Yara;<br>Almeida, 2005             |
| Rice         | <i>Oryza sativa</i> L.           | leaves and roots                    | <i>Chaetomium globosum</i> ,<br><i>Penicillium chrysogenum</i> , <i>Fusarium</i><br><i>oxysporum</i> and <i>Cladosporium</i><br><i>cladosporioides</i>  | Naik; Shashikala;<br>Krishnamurthy,<br>2009 |
| Rubber trees | <i>Hevea brasiliensis</i><br>L.  | leaves and stems                    | <i>Penicillium</i> , <i>Pestalotiopsis</i> and<br><i>Trichoderma</i>  | Gazis; Chaverri,<br>2010                    |
| Soybean      | <i>Glycine max</i> (L.)<br>Merr. | leaves and stems                    | (*1)  | Pimentel et al.,<br>2006                    |
|              |                                  | roots                               | (*1)  | Hamayun et al.,<br>2009a; 2009b             |
|              |                                  | leaves                              | <i>Colletotrichum</i> , <i>Cochliobolus</i> , <i>Fusarium</i><br>and <i>Xylaria</i>   | Leite et al., 2013                          |
|              |                                  | leaves and roots                    | <i>Ampelomyces</i> sp., <i>Cladosporium</i><br><i>cladosporioides</i> , <i>Colletotrichum</i><br><i>gloeosporioides</i> , <i>Diaporthe helianthi</i> ,<br><i>Guignardia mangiferae</i> , <i>Phoma</i> sp.,<br><i>Fusarium oxysporum</i> , <i>Fusarium solani</i><br>and <i>Fusarium</i> sp. | Fernandes et al.,<br>2015                   |
| Sugarcane    | <i>Saccharum</i> spp.            | leaves                              | Ascomycota phylum   | Stuart et al., 2010                         |
|              |                                  | leaves                              | (*1)  | Leme et al., 2013                           |
| Wheat        | <i>Triticum aestivum</i><br>L.   | leaves, stems,<br>glumes and grains | <i>Alternaria alternata</i> , <i>Cladosporium</i><br><i>herbarum</i> , <i>Epicoccum nigrum</i> ,<br><i>Cryptococcus</i> sp., <i>Rhodotorula rubra</i> ,<br><i>Penicillium</i> sp., and <i>Fusarium</i><br><i>graminearum</i>  | Larran et al, 2007                          |

(\*1): Unavailable or unclear information in reference.

Analyzing the data displayed in Table 1, spatial and seasonal fluctuations in endophytic fungi are evident. For example, an endophytic fungus may or may not occur in a given plant species, depending on the cultivation locality, environmental conditions and the growth stage of the plant. Moreover, same species of endophytes have been isolated from different host species (GAO et al., 2005; MUSSI-DIAS et al., 2012).

Endophytic fungi have been encountered in the leaves, fruit, branches and other parts of the cocoa plant (*T. cacao*) (RUBINI et al., 2005; MEJÍA et al., 2008; HANADA et al., 2010). The most frequent taxa are not the same for all plant organs.

However, there are taxa in common, the genus *Fusarium*, which Rubini et al. (2005) and Hanada et al. (2010) found to be dominant and Mejia et al. (2008) also detected as endophytic in the cocoa plant. Moreover, species of the genus *Fusarium* have been reported to be members of the endophytic fungal community of other crops, such as the cowpea (RODRIGUES; MENEZES, 2002), coffee (VEGA et al., 2010) and common bean (GONZAGA et al., 2014). Fungi from the genera *Colletotrichum*, *Epicoccum*, *Phomopsis* and *Xylaria* are also frequently reported endophytic taxa associated with economically important plants (ARAÚJO et al., 2001; SANTAMARÍA; BAYMAN, 2005; LEITE et al., 2013; TALONTSI et al., 2013; GONZAGA et al., 2014; SANTOS et al., 2016).

### ENDOPHYTIC FUNGI PROVIDE BENEFITS TO THE HOST

Through interactions with the host plant, endophytic fungi play several beneficial roles that contribute to the health of the host and either directly or indirectly lead to an increase in plant productivity, such as protection against diseases caused by pathogenic microorganisms, protection against herbivory, the promotion of plant growth and the production of secondary metabolites.

#### A) Protection against diseases caused by pathogenic microorganisms

This aspect is a major benefit of endophytic fungi, which are capable of protecting the plant against pathogens through different strategies, such as competition with pathogens for colonization sites and nutrients, the production of antibiotics, the induction of resistance in the host plant and other mechanisms (AZEVEDO et al., 2000; STROBEL et al., 2001; ARNOLD et al., 2003; CLARKE et al., 2006; OWNLEY et al., 2010).

#### B) Protection against herbivory

Another activity that has been attributed to endophytic fungi is the protection of plants against attacks from herbivorous insects through the production of toxins (Raps and Vidal, 1998). Investigations have shown that endophytic fungi can influence the preference and performance of herbivorous insects, which minimizes the harm caused to the plant (RAPS; VIDAL, 1998; MEISTER et al., 2006; OKI et al., 2009; CRAWFORD et al., 2010; GANGE et al., 2012).

#### C) Plant growth promotion

Endophytic fungi can promote the growth of the host plant through the synthesis of phytohormones and/or by increasing plant tolerance to abiotic stress (PEIXOTO NETO et al., 2002; REDMAN et al., 2002; BAE et al., 2009; HAMAYUN et al., 2009b; KHAN et al., 2015). This ability to stimulate plant growth is extremely important and can be explored in agricultural practices. Indeed, the use of plant growth-promoting microorganisms is an option to help modern agriculture face the challenges of increasing crop production and ensuring sustainability (LUZ et al., 2006).

Studies involving the use of endophytic fungi for the promotion of plants of economic interest have achieved promising results. Examples include the work done with corn (*Zea mays* L.), tobacco (*Nicotiana tabacum* L.) (VARMA et al., 1999), yellow passion fruit (*Passiflora edulis* Sims. f. sp. *flavicarpa* Deg.) (LUZ et al., 2006), cocoa (*T. cacao* L.) (BAE et al., 2009) and soybean (*G. max* (L.) Merr.) (HAMAYUN et al., 2009a; 2009b; KHAN et al., 2011). *Piriformospora indica*, which is a basidiomycete that endophytically colonizes the roots of numerous plants, is among the species of

endophytic fungi most commonly used in plant growth promotion (VARMA et al., 1999; PEIXOTO NETO et al., 2002; KUMAR et al., 2011).

#### *D) Production of secondary metabolites*

Endophytic fungi produce a wide variety of secondary metabolites, which, unlike primary metabolites, play an important role in the physiological processes of microorganisms (STROBEL; DAISY, 2003; SURYANARAYANAN et al., 2009; RÖNSBERG et al., 2013). These compounds are produced for specific reasons, such as social interactions or predation, and are therefore related to the ecology of the producing organisms (BRAGA et al., 1999; CONTI et al., 2012). Secondary metabolites produced by endophytic fungi are also a potential source of novel bioactive natural products that may have applications in different fields of study (STROBEL; DAISY, 2003; SURYANARAYANAN et al., 2009; KUMAR; KAUSHIK, 2012; RÖNSBERG et al., 2013).

### **BIOTECHNOLOGICAL APPLICATIONS OF ENDOPHYTIC FUNGI**

The two main biotechnological applications of endophytic fungi are the production of secondary metabolites of economic interest and the biological control of pathogens, which are the focus of the discussion in this review.

#### *A) Secondary metabolites of biotechnological interest*

In the context of the prospecting of natural products, microorganisms are considered excellent producers of chemicals with desirable bioactive properties and substantial potential in the pharmaceutical industry (SURYANARAYANAN et al., 2009; CONTI et al., 2012; KUMAR; KAUSHIK, 2012). Indeed, fungi produce a quantity of chemicals currently used in medicine as antimicrobial agents and are producers of different substances of economic interest, such as enzymes, vitamins, amino acids and steroids (BRAGA et al., 1999; CAFÊU et al., 2005; MARINHO et al., 2009; SURYANARAYANAN et al., 2012; BARA et al., 2013).

Endophytic fungi stand out among the microorganisms that have been prospected in search of natural products with biotechnological applications. The secondary metabolites produced by these fungi have been the focus of several studies and represent a promising source for the bioprospecting of novel molecules with potential applications in both the pharmaceutical industry as well as agricultural activities in the form of crop protection (STROBEL; DAISY, 2003; AMARAL; RODRIGUES-FILHO, 2010; KUMAR; Kaushik, 2013; TALONTSI et al., 2013).

As endophytic fungi have ubiquitous distribution (RODRIGUEZ et al., 2009), there is an excellent chance of finding new species in plants that make up different ecosystems (STROBEL; DAISY, 2003). Thus, there is a range of metabolites to be explored from the biotechnological standpoint.

The most striking account of metabolite production by endophytic fungi with bioactive properties is the cancer medication taxol, which was originally produced from the yew tree (*Taxus brevifolia*). The production of this compound by an endophytic fungus was first demonstrated in studies by Stierle et al. (1993), in which the authors demonstrated that *Taxomyces andreanea* found in the yew tree was able to produce taxol *in vitro* (STIERLE et al., 1993). Subsequent studies have also shown that different species of endophytic fungi are also able to produce taxol, such as *Pestalotiopsis microspora* isolated from *Taxus wallachiana* (Strobel et al., 1996), *Pestalotiopsis terminaliae* isolated from *Terminalia arjuna* (GANGADEVI; MUTHUMARY, 2009), *Glocladium* sp. isolated from *Taxus baccata* (SREEKANTH et al., 2009) and *Guignardia mangiferae* isolated from *Taxus media* (XIONG et al., 2013).

The discovery that numerous endophytic fungi can produce taxol led to the emergence of the possibility of using a novel, more efficient, less expensive process to produce this important drug (PEIXOTO NETO et al., 2002). The laboratory cultivation of fungi does not require the extraction of the compound from *Taxus brevifolia* or other producing plants (PEIXOTO NETO et al., 2002; SREEKANTH et al., 2009; XIONG et al., 2013). This is one of the advantages of the exploration of secondary metabolites of a fungal origin in relation to other sources, as microorganisms can be grown in large-scale fermenters, which eliminates the harm to the ecosystem that occurs with the removal of plants from natural areas and minimizes the danger of the local extinction of plant species that are collected for the extraction of medicinal products (PEIXOTO NETO et al., 2002; TAKAHASHI; LUCAS, 2008).

In addition to taxol, the scientific literature reports the discovery of secondary metabolites produced by endophytic fungi with activity of biotechnological interest, such as the inhibition of pathogenic organisms (MOMESSO et al., 2008; SILVA et al., 2010; BUDHIRAJA et al., 2013). The following are examples of such metabolites: phomopsichalasin produced by *Phomopsis* sp., which has antibacterial activity (HORN et al., 1995); cryptocandin produced by *Cryptosporiopsis* cf. *quercina*, which has antifungal activity (STROBEL et al., 1999); cercosporin produced by *Mycosphaerella* sp., which has antiparasitic action (MORENO et al., 2011); and cytochalasins produced by *Chaetomium globosum* and *Xylaria* sp., which have diverse biological activities, including cytotoxic action (MOMESSO et al., 2008; SILVA et al., 2010).

As with endophytes of medicinal plants, the biological activity of endophytic fungi of crops, such as coffee (*Coffea arabica* L.) (SETTE et al., 2006; FERNANDES et al., 2009), rice (*Oryza sativa* L.) (NAIK et al., 2009), sugarcane (*Saccharum officinarum* L.) (FÁVARO et al., 2012), cocoa (*T. cacao* L.) (TALONTSI et al., 2013) and wheat (*Triticum durum* Desf.) (SADRATI et al., 2013), has also been investigated.

### *B) Endophytic fungi as biological control agents of phytopathogens*

The biological control of phytopathogens involves the use of microorganisms that reduce the activity or survival of disease-causing agents in plants (OWNLEY et al., 2010). The most commonly described biocontrol mechanisms are antibiosis, competition for space, iron and other nutrients, parasitism and the induction of resistance in the host (MARIANO et al., 2004; BAILEY et al., 2008; CAO et al., 2009; OWNLEY et al., 2010).

Endophytic fungi have been studied with regard to the ability to act as inhibitors of other microorganisms for application as biocontrol agents as an alternative to chemical control. These fungi can be used either directly, in which the organism is applied live and acts as an antagonist, or indirectly, through the use of metabolites (LAZZARETTI; BETTIOL, 1997; GRIGOLETTI JÚNIOR et al., 2000; MORANDI; BETTIOL, 2009).

Hanada et al. (2010) conducted field experiments to assess the biocontrol properties of endophytic fungi isolated from cocoa (*T. cacao* L.) and cupuassu (*T. grandiflorum* (Willd. ex Spreng.) K. Schum.) against *Phytophthora palmivora*, the causal agent of cocoa black-pod rot disease, which is one of the most important pathogens in cocoa-producing areas throughout the world. Fungal species of the genera *Pestalotiopsis*, *Curvularia*, *Tolyopcladium* and *Fusarium* have demonstrated the greatest activity against this pathogen (HANADA et al., 2010). Rubini et al. (2005), Bailey et al. (2008) and Mejía et al. (2008) also evaluated the potential use of endophytic fungi isolated from cocoa (*T. cacao* L.) for the biological control of phytopathogens.

Rocha et al. (2011) analyzed the potential application of endophytic fungi from rubber trees (*Hevea brasiliensis* L.) for the biological control of the pathogen *Microcyclus ulei*, which causes damage to rubber tree crops. Garoé et al. (2012) assessed the potential use of endophytic fungi from grapevine (*Vitis vinifera* L.) for the biological control of *Botrytis cinerea*, which affects grape crops. Similar studies should be conducted to assist in the development of sustainable strategies for the control of diseases that affect crops and lead to huge financial losses.

### CLOSING REMARKS

The findings of the present review of the literature underscore the importance of endophytic fungi to host plants and the need for further studies on the isolation and identification of these organisms as well as the prospecting of secondary metabolites of biotechnological interest.

### AUTHORS CONTRIBUTION

The biologist Beatriz dos Santos Souza, from UFOPA, carried out the selection of bibliographic data and contributed to the preparation of the manuscript. The Professor MSc. Taidés Tavares dos Santos, from UFT, planned and guided the study, as well as approved the final proof of the manuscript.

### CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest.

### FUNDING

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

### REFERENCES

- ALMEIDA, C.V.; YARA, R.; ALMEIDA, M. Fungos endofíticos isolados de ápices caulinares de pupunheira cultivada *in vivo* e *in vitro*. **Pesquisa Agropecuária Brasileira**, v. 40, n. 5, p. 467–470, 2005.
- AMARAL, L.S.; RODRIGUES-FILHO, E. Two novel eremophilane sesquiterpenes from an endophytic Xylariaceae fungus isolated from leaves of *Cupressus lusitanica*. **Journal of the Brazilian Chemical Society**, v. 21, n. 8, p. 1446–1450, 2010. DOI:10.1590/S0103-50532010000800006
- ARAÚJO, W.L.; MACCHERONI, W.; AGUILAR-VILDOSO, C.I.; BARROSO, P.A.; SARIDAKIS, H.O.; AZEVEDO, J.L. Variability and interactions between endophytic bacteria and fungi isolated from leaf tissues of citrus rootstocks. **Canadian Journal of Microbiology**, v. 47, n. 3, p. 229–236, 2001. DOI: 10.1139/w00-146
- ARNOLD, A.E.; MEJIA, L.C.; KYLLO, D.; ROJAS, E.I.; MAYNARD, Z.; ROBBINS, N.; HERRE, E.A. Fungal endophytes limit pathogen damage in a tropical tree. **Proceedings of the National Academy of Sciences (PNAS)**, v. 100, n. 26, p. 15649–15654, 2003. DOI: 10.1073/pnas.2533483100
- ARNOLD, A.E.; MAYNARD, Z.; GILBERT, G.S. Fungal endophytes in dicotyledonous neotropical trees: patterns of abundance and diversity. **Mycological Research**, v.105, n.12, p.1502–1507, 2001. DOI:10.1017/S0953756201004956



- AZEVEDO, J.L.; MACCHERONI JUNIOR, W.; PEREIRA, J.O.; ARAÚJO, W.L. Endophytic microorganisms: a review on insect control and recent advances on tropical plants. **Electronic Journal of Biotechnology**, v. 3, n. 1, 40-65, 2000. DOI: [10.4067/S0717-34582000000100004](https://doi.org/10.4067/S0717-34582000000100004)
- BAE, H.; SICHER, R.C.; KIM, M.S.; KIM, S.H.; STREM, M.D.; MELNICK, R.L.; BAILEY, B.A. The beneficial endophyte *Trichoderma hamatum* isolate DIS 219b promotes growth and delays the onset of the drought response in *Theobroma cacao*. **Journal of Experimental Botany**, v. 60, n. 11, p. 3279–3295, 2009. DOI: [10.1093/jxb/erp165](https://doi.org/10.1093/jxb/erp165)
- BAILEY, B.A.; BAE, H.; STREM, M.D.; CROZIER, J.; THOMAS, S.E.; SAMUELS, G.J.; VINYARD, B.T.; HOLMES, K.A. Antibiosis, mycoparasitism, and colonization success for endophytic *Trichoderma* isolates with biological control potential in *Theobroma cacao*. **Biological Control**, v.46, p.24-35, 2008. DOI: [10.1016/j.biocontrol.2008.01.003](https://doi.org/10.1016/j.biocontrol.2008.01.003)
- BARA, R.; ALY, A.H.; WRAY, V.; LIN, W.; PROKSCH, P.; DEBBAB, A. Talaromins A and B, new cyclic peptides from the endophytic fungus *Talaromyces wortmannii*. **Tetrahedron Letters**, v. 54, n. 13, p. 1686-1689, 2013. DOI: [10.1016/j.tetlet.2013.01.064](https://doi.org/10.1016/j.tetlet.2013.01.064)
- BRAGA, G.U.L.; DESTÉFANO, R.H.R.; MESSIAS, C.L. Protease production during growth and autolysis of submerged *Metarhizium anisopliae* cultures. **Revista de Microbiologia**, v. 30, n. 2, p. 107-113, 1999. DOI: [10.1590/S0001-37141999000200004](https://doi.org/10.1590/S0001-37141999000200004)
- BUDHIRAJA, A.; NEPALI, K.; SAPRA, S.; GUPTA, S.; KUMAR, S.; DHAR, K.L. Bioactive metabolites from an endophytic fungus of *Aspergillus* species isolated from seeds of *Gloriosa superba* Linn. **Medicinal Chemistry Research**, v. 22, n. 1, p. 323–329, 2013. DOI: [10.1007/s00044-012-0032-z](https://doi.org/10.1007/s00044-012-0032-z)
- CABRAL, D.; STONE, J.K.; CARROLL, G.C. The internal mycobiota of *Juncus* spp.: microscopic and cultural observations of infection patterns. **Mycological Research**, v. 97, n. 3, p. 367-376, 1993. DOI: [10.1016/S0953-7562\(09\)81140-4](https://doi.org/10.1016/S0953-7562(09)81140-4)
- CAFÊU, M.C.; SILVA, G.H.; TELES, H.L.; BOLZANI, V.S.; ARAÚJO, A.R.; YOUNG, M.C.M.; PFENNING, L.H. Substâncias antifúngicas de *Xylaria* sp., um fungo endofítico isolado de *Palicourea marcgravii* (Rubiaceae). **Química Nova**, v.28, n.6, p.991-995, 2005.
- CAMPOS, F.F.; SALES JÚNIOR, P.A.; ROMANHA, A.J.; ARAÚJO, M.S.S.; SIQUEIRA, E.P.; RESENDE, J.M.R.; ALVES, T.M.A.; MARTINS-FILHO, A.O.; SANTOS, V.L.; ROSA, C.A.; ZANI, C.L.; COSTA, B.B. Bioactive endophytic fungi isolated from *Caesalpinia echinata* Lam. (Brazilwood) and identification of beauvericin as atryanocidal metabolite from *Fusarium* sp. **Memórias do Instituto Oswaldo Cruz**, v. 110, p. 65-74, 2015. DOI: [10.1590/0074-02760140243](https://doi.org/10.1590/0074-02760140243)
- CAO, R.; LIU, X.; GAO, K.; MENDGEN, K.; KANG, Z.; GAO, J.; DAI, Y.; WANG, X. Mycoparasitism of endophytic fungi isolated from reed on soilborne phytopathogenic fungi and production of cell wall-degrading enzymes *in vitro*. **Current Microbiology**, v. 59, p. 584–592, 2009. DOI: [10.1007/s00284-009-9477-9](https://doi.org/10.1007/s00284-009-9477-9)
- CHEN, Z.; CHEN, H.P.; LI, Y.; FENG, T.; LIU, J.K. Cytochalasins from cultures of endophytic fungus *Phoma multirostrata* EA-12. **The Journal of Antibiotics**, v. 68, p. 23-16, 2015. DOI: [10.1038/ja.2014.87](https://doi.org/10.1038/ja.2014.87)
- CLARKE, B.B.; WHITE JR, J.F.; HURLEY, R.H.; TORRES, M.S.; SUN, S.; HUFF, D.R. Endophyte-mediated suppression of dollar spot disease in fine fescues. **Plant Disease**, v.90, p.994–998, 2006. DOI: [10.1094/PD-90-0994](https://doi.org/10.1094/PD-90-0994)
- CONTI, R.; GUIMARÃES, D.O.; PUPO, M.T. Aprendendo com as interações da natureza: microrganismos simbiotes como fontes de produtos naturais bioativos. **Ciência e Cultura**, v.64, n.3, p.43-47, 2012. DOI: [10.21800/S0009-67252012000300014](https://doi.org/10.21800/S0009-67252012000300014)
- CRAWFORD, K.M.; LAND, J.M.; RUDGERS, J.A. Fungal endophytes of native grasses decrease insect herbivore preference and performance. **Oecologia**, v.164, n.2, p.431–444, 2010. DOI: [10.1007/s00442-010-1685-2](https://doi.org/10.1007/s00442-010-1685-2)

CROZIER, J.; THOMAS, S. E.; AIME, M.C.; EVANS, H.C.; HOLMES, K.A. Molecular characterization of fungal endophytic morphospecies isolated from stems and pods of *Theobroma cacao*. **Plant Pathology**, v. 55, n. 6, p. 783–791, 2006. DOI: [10.1111/j.1365-3059.2006.01446.x](https://doi.org/10.1111/j.1365-3059.2006.01446.x)

FÁVARO, L.C.L.; SEBASTIANES, F.L.S.; ARAÚJO, W.L. *Epicoccum nigrum* P16, a sugarcane endophyte, produces antifungal compounds and induces root growth. **Plos One**, v.7, n.6, p.1-10, 2012. DOI:[10.1371/journal.pone.0036826](https://doi.org/10.1371/journal.pone.0036826)

FERNANDES, M.R.V.; SILVA, T.A.C.; PFENNING, L.H.; COSTA-NETO, C.M.; HEINRICH, T.A.; ALENCAR, S.M.; LIMA, M.A.; IKEGAKI, M. Biological activities of the fermentation extract of the endophytic fungus *Alternaria alternata* isolated from *Coffea arabica* L. **Brazilian Journal of Pharmaceutical Sciences**, v. 45, n. 4, p. 677-685, 2009. DOI:[10.1590/S1984-82502009000400010](https://doi.org/10.1590/S1984-82502009000400010)

FERNANDES, E.G.; PEREIRA, O.L.; SILVA, C.C.; BENTO, C.B.P.; QUEIROZ, M.V. Diversity of endophytic fungi in *Glycine max*. **Microbiological Research**, v. 181, p. 84–92, 2015. DOI: [10.1016/j.micres.2015.05.010](https://doi.org/10.1016/j.micres.2015.05.010)

FERREIRA, M.C.; VIEIRA, M.L.A.; ZANI, C.Z.; ALVES, T.M.A.; SALES JUNIOR, P.A.; MURTA, S.M.F.; ROMANHA, A.J.; GIL, L.H.V.G.; CARVALHO, A.G.O.; ZILLI, J.E.; VITAL, M.J.S.; ROSA, C.A.; ROSA, L.H. Molecular phylogeny, diversity, symbiosis and discover of bioactive compounds of endophytic fungi associated with the medicinal Amazonian plant *Carapa guianensis* Aublet (Meliaceae). **Biochemical Systematics and Ecology**, v. 59, p. 36-44, 2015. DOI: [10.1016/j.bse.2014.12.017](https://doi.org/10.1016/j.bse.2014.12.017)

FISHER, P.J.; PETRINI, O.; SCOTT, H.M.L. The distribution of some fungal and bacterial endophytes in Maize (*Zea mays* L). **New Phytologist**, v. 122, p. 299-305, 1992. DOI: [10.1111/j.1469-8137.1992.tb04234.x](https://doi.org/10.1111/j.1469-8137.1992.tb04234.x)

GANGADEVI, V.; MUTHUMARY, J. Taxol production by *Pestalotiopsis terminaliae*, an endophytic fungus of *Terminalia arjuna* (arjun tree). **Biotechnology and Applied Biochemistry**, v.52, p.9–15, 2009. DOI:[10.1042/BA20070243](https://doi.org/10.1042/BA20070243)

GANGE, A.C.; ESCHEN, R.; WEARN, J.A.; THAWER, A.; SUTTON, B.C. Differential effects of foliar endophytic fungi on insect herbivores attacking a herbaceous plant. **Oecologia**, v. 168, n. 4, p. 1023–1031, 2012. DOI:[10.1007/s00442-011-2151-5](https://doi.org/10.1007/s00442-011-2151-5)

GAO, F.; DAI, C.; LIU, X. Mechanisms of fungal endophytes in plant protection against pathogens. **African Journal of Microbiology Research**, v. 4, n. 13, p. 1346–1351, 2010.

GAO, X.X.; ZHOU, H.; XU, D.-Y.; YU, C.-H.; CHEN, Y.-Q.; QU, L.-H. High diversity of endophytic fungi from the pharmaceutical plant, *Heterosmilax japonica* Kunth revealed by cultivation-independent approach. **FEMS Microbiology Letters**, v. 249, p. 255-266, 2005. DOI: [10.1016/j.femsle.2005.06.017](https://doi.org/10.1016/j.femsle.2005.06.017)

GAROÉ, N.; CABRERA, R.; LISBEL, B.R.; EVELYN, D.S.; ANDREEA, C.; NÉLIDA, B. Endophytic fungi from *Vitis vinifera* L. isolated in Canary Islands and Azores as potential biocontrol agents of *Botrytis cinerea* Pers.: Fr. **Journal of Horticulture, Forestry and Biotechnology**, v. 16, n. 1, p. 1–6., 2012.

GAZIS, R.; CHAVERRI, P. Diversity of fungal endophytes in leaves and stems of wild rubber trees (*Hevea brasiliensis*) in Peru. **Fungal Ecology**, v. 3, n. 3, p. 240–254, 2010. DOI: [10.1016/j.funeco.2009.12.001](https://doi.org/10.1016/j.funeco.2009.12.001)

GONZAGA, L.L.; COSTA, L.E.; SANTOS, T.T.; ARAÚJO, E.F.; QUEIROZ, M.V. Endophytic fungi from the genus *Colletotrichum* are abundant in the *Phaseolus vulgaris* and have high genetic diversity. **Journal of Applied Microbiology**, v. 118, p. 485–496, 2014. DOI: [10.1111/jam.12696](https://doi.org/10.1111/jam.12696)

GRIGOLETTI JUNIOR, A.; SANTOS, A.F.; AUER, C.G. Perspectivas do uso do controle biológico contra doenças florestais. **Floresta**, v. 30, p. 155-165, 2000.

HAMAYUN, M.; KHAN, S.; AHMAD, N.; TANG, D.S.; KANG, S.M.; NA, C.I.; SOHN, E.Y.; HWANG, Y.H.; SHIN, D.H.; LEE, B.H.; KIM, J.G.; LEE, I.J. *Cladosporium sphaerospermum* as a new plant growth-promoting endophyte from the roots of *Glycine max* (L.). Merr. **World Journal Microbiology and Biotechnology**, v. 25, n. 4, p. 627-632, 2009a. DOI: [10.1007/s11274-009-9982-9](https://doi.org/10.1007/s11274-009-9982-9)

- HAMAYUN, M.; KHAN, S.A.; KIM, H.Y.; CHAUDHARY, M.F.; HWANG, Y.H.; SHIN, D.H.; KIM, I.K.; LEE, B.H.; LEE, I.J. Gibberellin production and plant growth enhancement by newly isolated strain of *Scolecobasidium tshawytschae*. **Journal of Microbiology and Biotechnology**, v. 19, n. 6, p. 560-565, 2009b.
- HANADA, R.E.; JORGE SOUZA, T.; POMELLA, A.W.V.; HEBBAR, K.P.; PEREIRA, J.O.; ISMAIEL, A.; SAMUELS, G.J. *Trichoderma martiale* sp. nov., a new endophyte from sapwood of *Theobroma cacao* with a potential for biological control. **Mycological Research**, v. 112, p. 1335–1343, 2008. DOI: [10.1016/j.mycres.2008.06.022](https://doi.org/10.1016/j.mycres.2008.06.022)
- HANADA, R.E.; POMELLA, A.W.V.; COSTA, H.S.; BEZERRA, J.L.; LOGUERCIO, L.L.; PEREIRA, J.O. Endophytic fungal diversity in *Theobroma cacao* (cacao) and *T. grandiflorum* (cupuaçu) trees and their potential for growth promotion and biocontrol of black-pod disease. **Fungal Biology**, v. 114, p. 901–910, 2010. DOI: [10.1016/j.funbio.2010.08.006](https://doi.org/10.1016/j.funbio.2010.08.006)
- HORN, W.S.; SIMMONDS, M.S.J.; SCHWARTZ, R.E.; BLANEY, W.M. Phomopsichalasin, a novel antimicrobial agent from an endophytic *Phomopsis* sp. **Tetrahedron**, v. 51, n. 14, 3969–3978, 1995. DOI: [10.1016/0040-4020\(95\)00139-Y](https://doi.org/10.1016/0040-4020(95)00139-Y)
- HYDE, K.D.; SOYTONG, K. The fungal endophyte dilemma. **Fungal Diversity**, v. 33, p. 163-173, 2008.
- KHAN, A.L.; HAMAYUN, M.; AHMAD, N.; HUSSAIN, J.; KANG, S.M.; KIM, Y.H.; ADNAN, M.; TANG, D.S.; WAQAS, M.; RADHAKRISHNAN, R.; HWANG, Y.H.; LEE, I.J. Salinity Stress Resistance Offered by Endophytic Fungal Interaction Between *Penicillium minioluteum* LHL09 and Glycine max. L. **Journal of Microbiology and Biotechnology**, v. 21, p. 893–902, 2011. DOI: [10.4014/jmb.1103.03012](https://doi.org/10.4014/jmb.1103.03012)
- KHAN, A.R.; ULLAH, I.; WAQAS, M.; SHAHZAD, R.; HONG, S.-J.; PARK, G.-S.; JUNG, B.K.; LEE, I.-J.; SHIN, J.-H. Plant growth-promoting potential of endophytic fungi isolated from *Solanum nigrum* leaves. **World Journal of Microbiology and Biotechnology**, v. 31, n. 9, p. 1461–1466, 2015. DOI: [10.1007/s11274-015-1888-0](https://doi.org/10.1007/s11274-015-1888-0)
- KUMAR, S.; KAUSHIK, N. Metabolites of endophytic fungi as novel source of biofungicide: a review. **Phytochemistry Review**, v. 11, p. 507–522, 2012. DOI: [10.1007/s11101-013-9271-y](https://doi.org/10.1007/s11101-013-9271-y)
- KUMAR, S.; KAUSHIK, N. Endophytic Fungi Isolated from Oil-Seed Crop *Jatropha curcas* Produces Oil and Exhibit Antifungal Activity. **Plos One**, v. 8, n. 2, p. 1-8, 2013. DOI: [10.1371/journal.pone.0056202](https://doi.org/10.1371/journal.pone.0056202)
- KUMAR, V.; SAHAI, V.; BISARIA, V.S. High-density spore production of *Piriformospora indica*, a plant growth-promoting endophyte, by optimization of nutritional and cultural parameters. **Bioresource Technology**, v. 102, p. 3169–3175, 2011. DOI: [10.1016/j.biortech.2010.10.116](https://doi.org/10.1016/j.biortech.2010.10.116)
- KUMAR, S.; UPADHYAY, R.; AHARWAL, R.P.; SANDHU, S.S. Antibacterial activity of some isolated endophytic fungi from *Mentha viridis*. **International Journal of Applied Biology and Pharmaceutical Technology**, v. 7, n. 1, p. 239–249, 2016.
- LARRAN, S.; PERELLO, A.; SIMON, M.R.; MORENO, V. The endophytic fungi from wheat (*Triticum aestivum* L.). **World Journal Microbiology and Biotechnology**, v. 23, p. 565-572, 2007. DOI: [10.1007/s11274-006-9266-6](https://doi.org/10.1007/s11274-006-9266-6)
- LAZZARETTI, E.; BETTIOL, W. Tratamento de sementes de arroz, trigo, feijão e soja com um produto formulado à base de células e de metabólitos de *Bacillus subtilis*. **Scientia Agricola**, v. 54, p. 89-96, 1997. DOI: [10.1590/S0103-90161997000100013](https://doi.org/10.1590/S0103-90161997000100013)
- LEITE, T.S.; CNOSSEN-FASSONI, A.; PEREIRA, O.L.; MIZUBUTI, E.S.G.; ARAÚJO, E.F.; QUEIROZ, M.V. Novel and Highly Diverse Fungal Endophytes in Soybean Revealed by the Consortium of Two Different Techniques. **Journal of Microbiology**, v. 51, n. 1, p. 56–69, 2013. DOI: [10.1007/s12275-013-2356-x](https://doi.org/10.1007/s12275-013-2356-x)
- LEME, A.C.; BEVILAQUA, M.R.R.; RHODEN, S.A.; MANGOLIN, C.A.; MACHADO, M.F.P.S.; PAMPHILE, J.A. Molecular characterization of endophytes isolated from *Saccharum* spp based on esterase and ribosomal DNA (ITS1-5.8S-ITS2) analyses. **Genetics and Molecular Research**, v. 12, n. 3, p. 4095–105, 2013. DOI: [10.4238/2013](https://doi.org/10.4238/2013)

- LUZ, J.S.; SILVA, R.L.O.; SILVEIRA, E.B.; CAVALCANTE, U.M.T. Atividade enzimática de fungos endofíticos e efeito na promoção do crescimento de mudas de maracujazeiro-amarelo. **Revista Caatinga**, v. 19, n. 2, p. 128-134, 2006.
- MARIANO, R.L.R.; SILVEIRA, E.B.; ASSIS, S.M.P.; GOMES, A.M.A.; NASCIMENTO, A.R.P.; DONATO, V.M.T.S. Importância de bactérias promotoras de crescimento e de biocontrole de doenças de plantas para uma agricultura sustentável. **Anais da Academia Pernambucana de Ciência Agronômica**, v. 1, p. 89-111, 2004.
- MARINHO, A.M.R.; MARINHO, P.S.B.; RODRIGUES FILHO, E. Esteroides produzidos por *Penicillium herquei*, um fungo endofítico isolado dos frutos de *Melia azedarach* (Meliaceae). **Química Nova**, v. 32, n. 7, p. 1710-1712, 2009.
- MEISTER, B.; KRAUSS, J.; HÄRRI, S.A.; SCHNEIDER, M.V.; MULLER, C.B. Fungal endosymbionts affect aphid population size by reduction of adult life span and fecundity. **Basic and Applied Ecology**, v. 7, p. 244- 252, 2006. DOI: [10.1016/j.baae.2005.06.002](https://doi.org/10.1016/j.baae.2005.06.002)
- MEJÍA, L.C.; ROJAS, E.I.; MAYNARD, Z.; BAEL, S.V.; ARNOLD, A.E.; HEBBAR, P.; SAMUELS, G.J.; ROBBINS, N.; HERRE, E.A. Endophytic fungi as biocontrol agents of *Theobroma cacao* pathogens. **Biological Control**, v. 46, p. 4–14, 2008. DOI: [10.1016/j.biocontrol.2008.01.012](https://doi.org/10.1016/j.biocontrol.2008.01.012)
- MOMESSO, L.S.; KAWANO, C.Y.; RIBEIRO, P.H.; NOMIZO, A.; GOLDMAN, G.H.; PUPO, M.T. Chaetoglobosinas produzidas por *Chaetomium globosum*, fungo endofítico associado a *Viguiera robusta* Gardn. (Asteraceae). **Química Nova**, v. 31, n. 7, p. 1680-1685, 2008.
- MORANDI, M.A.B.; BETTIOL, W. Controle biológico de doenças de plantas no Brasil. In: BETTIOL, W.; MORANDI, M.A.B. (Ed.) **Biocontrole de doenças de plantas: uso e perspectivas**. Embrapa Meio Ambiente, 2009, 341 p.
- MORENO, E.; VARUGHESE, T.; SPADAFORA, C.; ARNOLD, E.; COLEY, P.D.; KURSAR, T.A.; GERWICK, W.H.; CUBILLA-RIOS, L. Chemical Constituents of the New Endophytic Fungus *Mycosphaerella* sp. nov. and Their Anti-Parasitic Activity. **Natural Product Communications**, v. 6, p. 835-840, 2011.
- MÜLLER, C.B.; KRAUSS, J. Symbiosis between grasses and asexual fungal endophytes. **Current Opinion in Plant Biology**, v. 8, n. 4, p. 450-456, 2005. DOI: [10.1016/j.pbi.2005.05.007](https://doi.org/10.1016/j.pbi.2005.05.007)
- MUSSI-DIAS, V.; ARAÚJO, A.C.O.; SILVEIRA, S.F.; ROCABADO, J.M.A.; ARAÚJO, K.L. Fungos endofíticos associados a plantas medicinais. **Revista Brasileira de Plantas Mediciniais**, v. 14, n. 2, p. 261-266, 2012.
- NAIK, B.S.; SHASHIKALA, J.; KRISHNAMURTHY, Y.L. Study on the diversity of endophytic communities from rice (*Oryza sativa* L.) and their antagonistic activities in vitro. **Microbiological Research**, v. 164, n. 3, p. 290–296, 2009. DOI: [10.1016/j.micres.2006.12.003](https://doi.org/10.1016/j.micres.2006.12.003)
- NISA, H.; KAMILI, A.N.; NAWCHOO, I.A.; SHAFI, S.; SHAMEEM, N.; BANDH, S.A. Fungal endophytes as prolific source of phytochemicals and other bioactive natural products: A review. **Microbial Pathogenesis**, v. 82, p. 50–59, 2015. DOI: [10.1016/j.micpath.2015.04.001](https://doi.org/10.1016/j.micpath.2015.04.001)
- OKI, Y.; SOARES, N.; BELMIRO, M.S.; CORRÊA JR., A.; FERNANDES, G.W. Influência dos fungos endofíticos sobre os herbívoros de *Baccharis dracunculifolia* (Asteraceae). **Neotropical Biology and Conservation**, v. 4, n. 2, p. 83-88, 2009.
- OROLE, O.O.; ADEJUMO, T.O. Bacterial and fungal endophytes associated with grains and roots of maize. **Journal of Ecology and The Natural Environment**, v. 3, p. 298-303, 2011.
- OWNLEY, B.H.; GWINN, K.D.; VEGA, F.E. Endophytic fungal entomopathogens with activity against plant pathogens: ecology and evolution. **BioControl**, v. 55, p. 113–128, 2010. DOI: [10.1007/s10526-009-9241-x](https://doi.org/10.1007/s10526-009-9241-x)
- PANCHER, M.; CEOL, M.; CORNEO, P.E.; LONGA, C.M.O.; YOUSAF, S.; PERTOT, I.; CAMPISANO, A. Fungal endophytic communities in grapevines (*Vitis vinifera* L.) respond to crop management. **Applied and Environmental Microbiology**, v. 78, n. 12, p. 4308–4317, 2012. DOI: [10.1128/AEM.07655-11](https://doi.org/10.1128/AEM.07655-11)

- PEIXOTO NETO, P.A.S.; AZEVEDO, J.L.; ARAÚJO, W.L. Microrganismos endofíticos. **Biotecnologia Ciência & Desenvolvimento**, v. 29, p. 62-77, 2002.
- PEREIRA, J.; VIEIRA, M.; AZEVEDO, J. Endophytic fungi from *Musa acuminata* and their reintroduction into axenic plants. **World Journal of Microbiology and Biotechnology**, v. 15, p. 37-40, 1999. DOI:10.1023/A:1008859823806
- PETRINI, O. Fungal endophytes of tree leaves. In: Andrews, J.H.; Hirano, S.S. (eds.) **Microbial ecology of leaves**. New York: Springer-Verlag, 1991, p. 179-197.
- PIMENTEL, I.C.; GLIENKE-BLANCO, C.; GABARDO, J.; STUART, R.M.; AZEVEDO, J.L. Identification and colonization of endophytic fungi from soybean (*Glycine max* (L.) merril) under different environmental conditions. **Brazilian Archives of Biology and Technology**, v. 49, n. 5, p. 705-711, 2006. DOI:10.1590/S1516-89132006000600003
- RAPS, A.; VIDAL, S. Indirect effects of an unspecialized endophytic fungus on specialized plant – herbivorous insect interactions. **Oecologia**, v. 114, p. 541-547, 1998. DOI: 10.1007/s004420050478
- REDMAN, R.S.; SHEEHAN, K.B.; STOUT, T.G.; RODRIGUEZ, R.J.; HENSON, J.M. Thermotolerance generated by plant/fungal symbiosis. **Science**, v. 298, p. 1581, 2002.
- REKHA, D.; SHIVANA, M.B. Diversity, antimicrobial and antioxidant activities of fungal endophytes in *Cynodon dactylon* (L.) Pers. and *Dactyloctenium aegyptium* (L.) P. Beauv. **International Journal of Current Microbiology and Applied Science**, v. 3, n. 8, p. 573-591, 2014.
- ROCHA, A.C.S.; GARCIA, D.; UETANABARO, A.P.T.; CARNEIRO, R.T.O.; ARAÚJO, I.S.; MATTOS, C.R.R.; GÓES-NETO, A. Foliar endophytic fungi from *Hevea brasiliensis* and their antagonism on *Microcyclus ulei*. **Fungal Diversity**, v. 47, p. 75-84, 2011. DOI:10.1007/s13225-010-0044-2
- RODRIGUES, A.A.C.; MENEZES, M. Detecção de fungos endofíticos em sementes de caupi provenientes de Serra Talhada e de Caruaru, Estado do Pernambuco. **Fitopatologia Brasileira**, v. 27, n. 5, p. 532-537, 2002.
- RODRIGUEZ, R.J.; WHITE JUNIOR, J.F.; ARNOLD, A.E.; REDMAN, R.S. Fungal endophytes: diversity and functional roles. **The New Phytologist**, v. 182, n. 2, p. 314-330, 2009. DOI:10.1111/j.1469-8137.2009.02773.x
- RÖNSBERG, D.; DEBBAB, A.; MÁNDI, A.; WRAY, V.; DAI, H.; KURTÁN, T.; PROKSCH, P.; ALY, A.H. Secondary metabolites from the endophytic fungus *Pestalotiopsis virgatula* isolated from the mangrove plant *Sonneratia caseolaris*. **Tetrahedron Letters**, v.54, n.25, p.3256-3259, 2013. DOI:10.1016/j.tetlet.2013.04.031
- RUBINI, M.R.; SILVA-RIBEIRO, R.T.; POMELLA, A.W.V.; MAKI, C.S.; ARAÚJO, W.L.; SANTOS, D.R.; AZEVEDO, J.L. Diversity of endophytic fungal community of cacao (*Theobroma cacao* L.) and biological control of *Crinipellis pernicioso*, causal agent of Witches' Broom Disease. **International Journal of Biological Sciences**, v. 1, p. 24-33, 2005.
- SADRATI, N.; DAOUD, H.; ZERROUG, A.; DAHAMNA, S.; BOUHARATI, S. Screening of antimicrobial and antioxidant secondary metabolites from endophytic fungi isolated from wheat (*Triticum durum*). **Journal of Plant Protection Research**, v. 53, n. 2, p. 128-136, 2013. DOI: 10.2478/jppr-2013-0019
- SANTAMARÍA, J.; BAYMAN, P. Fungal epiphytes and endophytes of coffee leaves (*Coffea arabica*). **Microbial Ecology**, v. 50, p. 1-8, 2005. DOI: 10.1007/s00248-004-0002-1
- SANTOS, T.T.; LEITE, T.S.; QUEIROZ, C.B.; ARAÚJO, E.F.; PEREIRA, O.L.; QUEIROZ, M.V. High genetic variability in endophytic fungi from the genus *Diaporthe* isolated from common bean (*Phaseolus vulgaris* L.) in Brazil. **Journal of Applied Microbiology**, v. 120, n. 2, p. 388-401, 2016. DOI: 10.1111/jam.12985
- SETTE, L.D.; PASSARINI, M.R.Z.; DELARMELENA, C.; SALATI, F.; DUARTE, M.C.T. Molecular characterization and antimicrobial activity of endophytic fungi from coffee plants. **World Journal Microbiology and Biotechnology**, v. 22, p. 1185-1195, 2006. DOI: 10.1007/s11274-006-9160-2

SILVA, G.H.; OLIVEIRA, C.M.; TELES, H.L.; BOLZANI, V.S.; ARAUJO, A.R.; PFENNING, L.H.; YOUNG, M.C.M.; COSTA-NETO, C.M.; HADDAD, R.; EBERLIN, M.N. Citocalasinas produzidas por *Xylaria* sp., um fungo endofítico de *Piper aduncum* (piperaceae). **Química Nova**, v. 33, n. 10, p. 2038-2041, 2010.

SOUZA, A.Q.L.; SOUZA, A.D.L.; ASTOLFI FILHO, S.; BELÉM PINHEIRO, M.L.; SARQUIS, M.I.M.; PEREIRA, J.O. Atividade antimicrobiana de fungos endofíticos isolados de plantas tóxicas da amazônia: *Palicourea longiflora* (aubl.) rich e *Strychnos cogens* bentham. **Acta Amazônica**, v. 34, n. 2, p. 185-195, 2004.

SREEKANTH, D.; SYED, A.; SARKAR, S.; SARKAR, D.; SANTHAKUMARI, B.; AHMAD, A.; KHAN, M.I. Production, purification, and characterization of Taxol and 10-DABIII from a new Endophytic Fungus *Gliocladium* sp. Isolated from the Indian Yew Tree, *Taxus baccata*. **Journal of Microbiology and Biotechnology**, v. 19, n. 11, p. 1342–1347, 2009. DOI: [10.4014/jmb.0904.04041](https://doi.org/10.4014/jmb.0904.04041)

STIERLE, A.; STROBEL, G.; STIERLE, D. Taxol and taxane production by taxomyces andreanae an endophytic fungus of Pacific yew. **Science**, v. 260, p. 214-216, 1993.

STROBEL, G.; DAISY, B. Bioprospecting for microbial endophytes and their natural products. **Microbiology and Molecular Biology Reviews**, v. 67, n. 4, p. 491-502, 2003. DOI: [10.1128/MMBR.67.4.491-502.2003](https://doi.org/10.1128/MMBR.67.4.491-502.2003)

STROBEL, G.; YANG, X.; SEARS, J.; KRAMER, R.; SIDHU, R.S.; HESS, W.M. Taxol from *Pestalotiopsis microspora*, an endophytic fungus of *Taxus wallachiana*. **Microbiology**, v. 142, p. 435-440, 1996. DOI:[10.1099/13500872-142-2-435](https://doi.org/10.1099/13500872-142-2-435)

STROBEL, G.A.; DIRKSE, E.; SEARS, J.; MARKWORTH, C. Volatile antimicrobials from *Muscodor albus*, a novel endophytic fungus. **Microbiology**, v. 147, p. 2943–2950, 2001. DOI: [10.1099/00221287-147-11-2943](https://doi.org/10.1099/00221287-147-11-2943)

STROBEL, G.A.; MILLER, R.V.; MARTINEZMILLER, C.; CONDRON, M.M.; TELOW, D.B.; HESS, W.M. Cryptocandin, a potent antimycotic from the endophytic fungus *Cryptosporiopsis* cf. *quercina*. **Microbiology**, v. 145, p. 1919-1926, 1999. DOI: [10.1099/13500872-145-8-1919](https://doi.org/10.1099/13500872-145-8-1919)

STUART, R.M.; ROMÃO, A.S.; PIZZIRANI-KLEINER, A.A.; AZEVEDO, J.L.; ARAÚJO, W.L. Culturable endophytic filamentous fungi from leaves of transgenic imidazolinone-tolerant sugarcane and its non-transgenic isolines. **Archives of Microbiology**, v.192, n.4, p.307-313, 2010. DOI: [10.1007/s00203-010-0557-9](https://doi.org/10.1007/s00203-010-0557-9)

SURYANARAYANAN, T.S.; THIRUNAVUKKARASU, N.; GOVINDARAJULU, M.B.; GOLAPAN, V. Fungal endophytes: an untapped source of biocatalysts. **Fungal Diversity**, v.54, n.1, p.19-30, 2012. DOI:[10.1007/s13225-012-0168-7](https://doi.org/10.1007/s13225-012-0168-7)

SURYANARAYANAN, T.S.; THIRUNAVUKKARASU, N.; GOVINDARAJULU, M.B.; SASSE, F.; JANSEN, R.; MURALI, T.S. Fungal endophytes and bioprospecting. **Fungal Biology Reviews**, v.23, p.9-19, 2009. DOI:[10.1016/j.fbr.2009.07.001](https://doi.org/10.1016/j.fbr.2009.07.001)

TAKAHASHI, J.A.; LUCAS, E.M.F. Ocorrência e diversidade estrutural de metabólitos fúngicos com atividade antibiótica. **Química Nova**, v.31, n.7, p.1807-1813, 2008.

TALONTSI, F.M.; DITTRICH, B.; SCHÜFFLER, A.; SUN, H.; LAATSCH, H. Epicoccolides: Antimicrobial and Antifungal Polyketides from an Endophytic Fungus *Epicoccum* sp. Associated with *Theobroma cacao*. **European Journal of Organic Chemistry**, v. 2013, n. 15, p. 3174–3180, 2013. DOI: [10.1002/ejoc.201300146](https://doi.org/10.1002/ejoc.201300146)

VARMA, A.; VERMA, S.; SUDAH, S.N.; FRANKEN, P. *Piriformospora indica*, a cultivable plant-growth promoting root endophyte. **Applied and Environmental Microbiology**, v.65, p.2741-2744, 1999.

VEGA, F.E.; SIMPKINS, A.; AIME, M.C.; POSADA, F.; PETERSON, S.W.; REHNER, S.A.; INFANTE, F.; CASTILLO, A.; ARNOLD, A.E. Fungal endophyte diversity in coffee plants from Colombia, Hawai'i, Mexico and Puerto Rico. **Fungal ecology**, v.3, p. 122-138, 2010. DOI: [10.1016/j.funeco.2009.07.002](https://doi.org/10.1016/j.funeco.2009.07.002)

XIONG, Z.Q.; YANG, Y.Y.; ZHAO, N.; WANG, Y. Diversity of endophytic fungi and screening of fungal paclitaxel producer from Anglojap yew, *Taxus x media*. **BMC Microbiology**, v.13, n.1, p.1-10, 2013. DOI:[10.1186/1471-2180-13-71](https://doi.org/10.1186/1471-2180-13-71)